

Fitting Dose Response Curve

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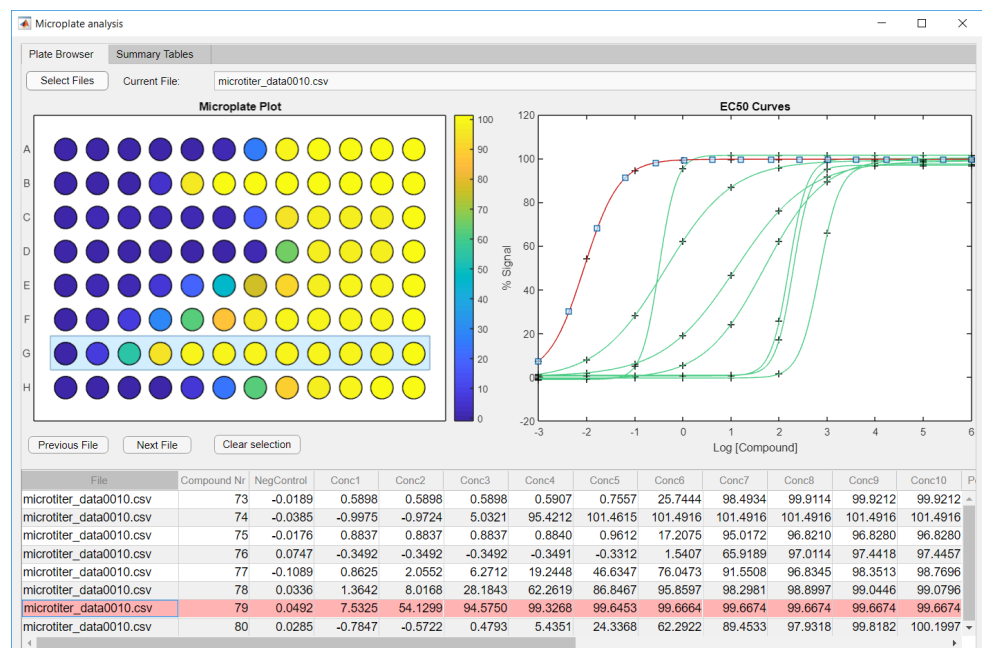
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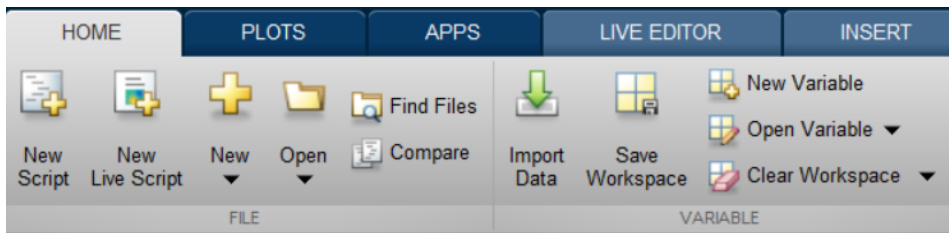
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This is an example of how MATLAB can be used to facilitate a typical workflow for an experimentalist who needs to automate analysis across many (hundreds) of experiments with instrument data organized in flat files, TXT, CSV files, or spreadsheets. In this example, we want to find the top leads from hundreds experiments. The data are in CSV files from a Dose-Response Experiment in a 96-well plate format. Each row of data corresponds to a serial dilution (middle 10 wells) of a given compound with a negative control (first column) and positive control (last column. The dose response readings in each microwell are in arbitrary units (e.g. fluorescence, luminescence, absorbance) ranging from ~0 to ~100(%). We fit the experimental data to a four parameter model and add a quality control metric drawn from the scientific literature. In the A_Microtiter_Plate_One file we prototyped our workflow. In this script we are calling that workflow in a loop. The final result is a table of compounds sorted in decreasing potency and filtered by a quality control metric. This also includes a report, and a spreadsheet file.



```
clear; close all; clc;
```

Step 1: Access Data in MATLAB



Use the **Import Data** option from the toolstrip to bring in your data for analysis or use the **readtable** function

```
data = readtable('data.csv')
```

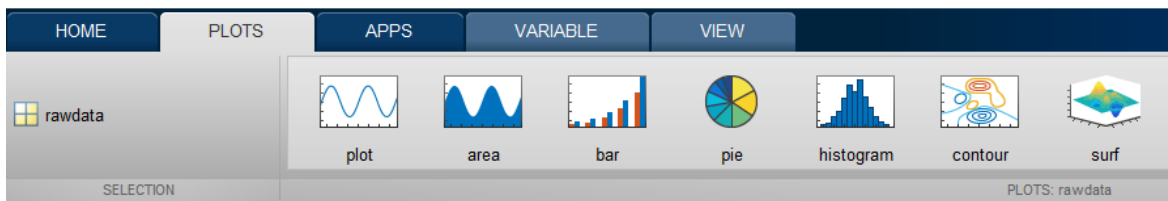
```
data = 8x12 table
```

...

	NegControl	Conc1	Conc2	Conc3	Conc4	Conc5	Conc6	Conc7
1	-0.0937	0.3389	9.9060	57.1390	92.1710	97.1720	97.6190	97.6570
2	-0.0336	-0.5264	-0.5264	-0.5264	-0.5264	-0.5264	-0.5263	-0.4762
3	-0.0137	-0.4983	3.2407	53.9460	99.3890	102.3700	102.4800	102.4800
4	0.0905	0.2136	0.2196	0.3525	3.2192	41.1440	94.8470	100.7800
5	-0.0775	-0.7785	1.7212	27.3410	85.9810	100.9600	102.2200	102.3100
6	0.0473	-0.0047	0.1516	1.9478	19.0260	73.4120	97.0630	99.8060
7	-0.0149	-0.3653	-0.3653	-0.3653	-0.3653	-0.3545	9.0751	100.0300
8	-0.0584	0.8869	0.8869	0.8870	0.9435	21.6330	101.4800	102.3300

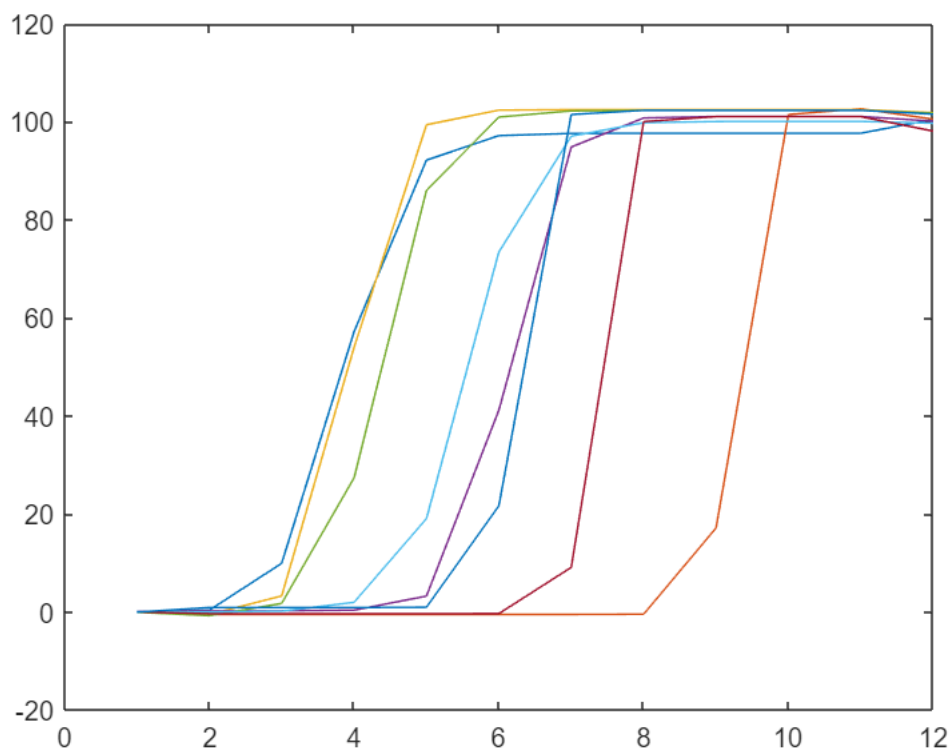
Step 2: Visualize the Data

When a variable in the workspace is selected, the **Plots Tab** automatically lights up with available visualizations that can accept that variable as input:



Let's try out a few of these visualizations and use the provided customization tools to adjust and automate custom data visualization

```
plot(data.Variables')
```



```
%bar3(data.Variables)
```

Step 3: Preprocess the Data

The data was imported into a **table**, one of several types of data containers. Tables are designed for columnar or tabular data, so accessing data can be done easily by name or by row and column numbers. The **table** also contains metadata accessible through properties.

```
data.Properties
```

```
ans =
```

```
TableProperties with properties:
```

```

    Description: ''
    UserData: []
    DimensionNames: {'Row' 'Variables'}
    VariableNames: {'NegControl' 'Conc1' 'Conc2' 'Conc3' 'Conc4' 'Conc5' 'Conc6' 'Conc7' 'Conc8' 'Conc9'}
    VariableDescriptions: {}
    VariableUnits: {}
    VariableContinuity: []
    RowNames: {}
    CustomProperties: No custom properties are set.
    Use addprop and rmprop to modify CustomProperties.
```

For example, let's assign row names to label each compound tested in the microtiter plate.

```
data.Properties.RowNames = cellstr("Compound" + (1:8))
```

data = 8x12 table

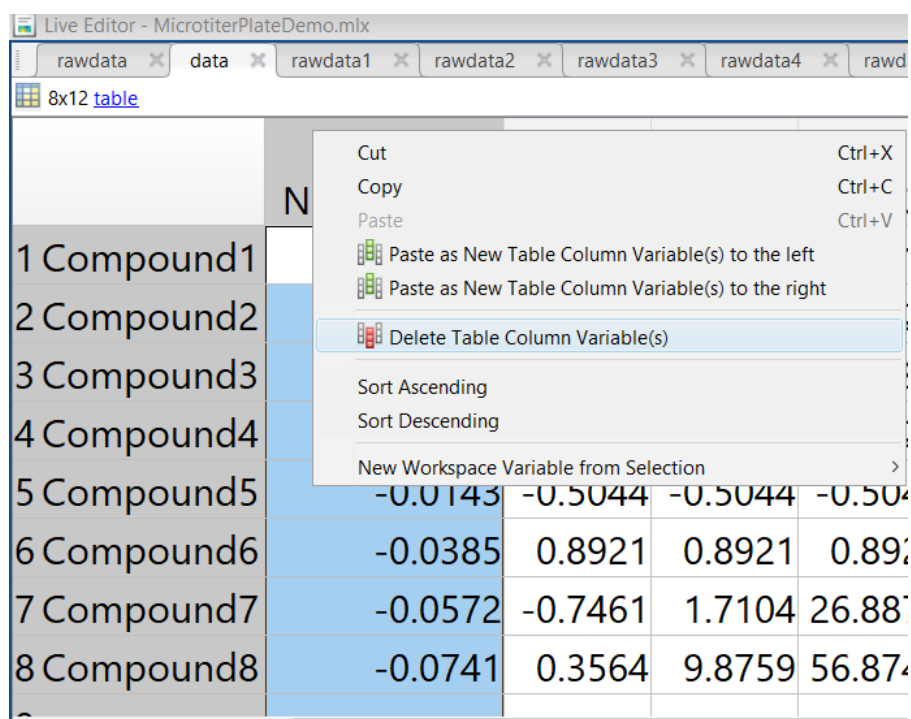
	NegControl	Conc1	Conc2	Conc3	Conc4	Conc5	Conc6	Conc7
1 Compound1	-0.0937	0.3389	9.9060	57.1390	92.1710	97.1720	97.6190	97.6570
2 Compound2	-0.0336	-0.5264	-0.5264	-0.5264	-0.5264	-0.5264	-0.5263	-0.4762
3 Compound3	-0.0137	-0.4983	3.2407	53.9460	99.3890	102.3700	102.4800	102.4800
4 Compound4	0.0905	0.2136	0.2196	0.3525	3.2192	41.1440	94.8470	100.7800
5 Compound5	-0.0775	-0.7785	1.7212	27.3410	85.9810	100.9600	102.2200	102.3100
6 Compound6	0.0473	-0.0047	0.1516	1.9478	19.0260	73.4120	97.0630	99.8060
7 Compound7	-0.0149	-0.3653	-0.3653	-0.3653	-0.3653	-0.3545	9.0751	100.0300
8 Compound8	-0.0584	0.8869	0.8869	0.8870	0.9435	21.6330	101.4800	102.3300

Before we can fit the data to our equation, we must first apply the positive and negative control measurements.

How do you do this in excel?

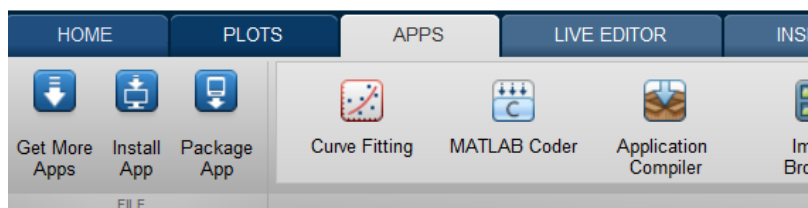
```
data.Variables = data.Variables - mean(data.NegControl);
data.Variables = data.Variables ./ data.PosControl*100;
```

Once this is done, we can remove the positive/negative control columns from our dataset. Try this from the **Variable Editor** and copy and paste the code below.



```
data = removevars(data, 'NegControl');
data = removevars(data, 'PosControl');
```

Step 4: Analyze the Data



MATLAB apps allow for easy and rapid prototyping while automatically generating the code for automation. Use the **Curve Fitting App** to fit each compound to the following equation:

$$R = \frac{\min - \max}{1 + (x/EC50)^{HillsSlope}} + \max$$

where:

$$1e-5 \leq EC50 \leq 1e7$$

$$0 \leq HillsSlope \leq 10$$

$$0 \leq \max \leq 120$$

$$-20 \leq \min \leq 5$$

and input concentrations range from 1nM to 1M and we visualize them as a sigmoidal curve:

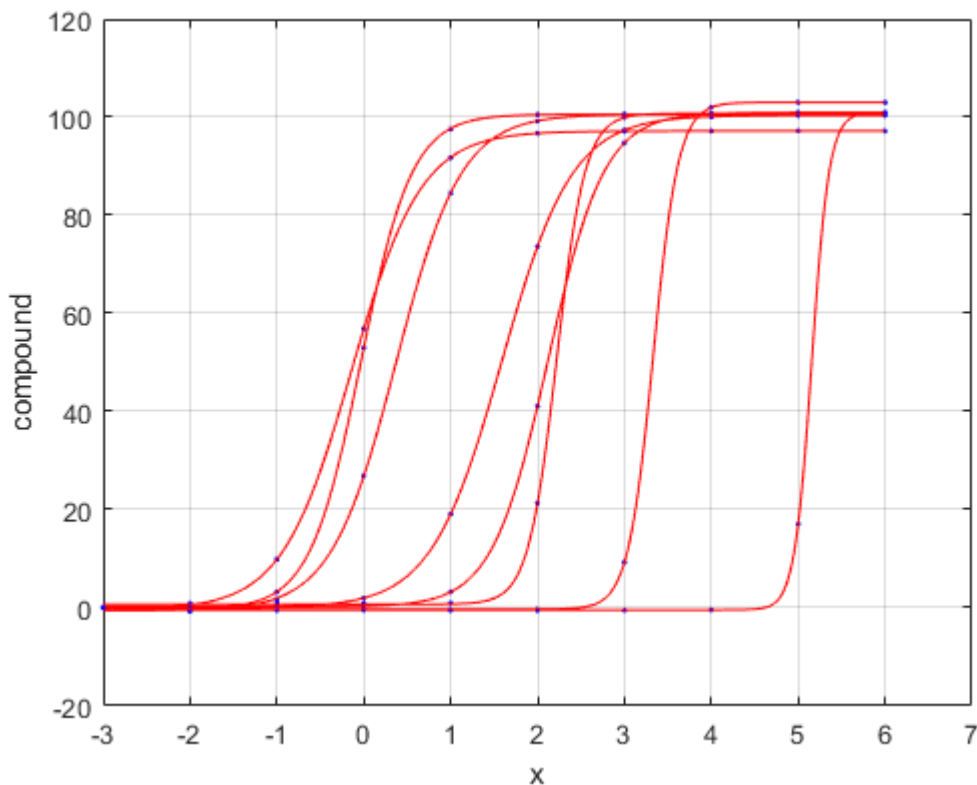
```
conc = [0.001, 0.01, 0.1, 1, 10, 100, 1000, 10000, 100000, 1000000];
x = log10(conc);
```

Once we've gone through the fitting for one compound, we can ask the app to generate the MATLAB code in order to repeat the process for the rest of the data.

Step 5: Automate Data Analysis

In order to repeat the same analysis process for the entire dataset, use a **for** loop. A loop is a type of programming construct that allows control over the flow a process. Loop control statements permit a particular sequence of commands to be repeated any number of times. Below, the **for** loop repeats the fitting process for the full data **table**.

```
clf
h = height(data);
EC50resultsmtx = zeros(h,1);
for ii = 1:h
    compound = data{ii,1:10};
    [fitresult, gof] = createMicrotiterFit(x, compound);
    EC50resultsmtx(ii,:) = [fitresult.EC50];
end
```



EC50resultsmtx

```
EC50resultsmtx = 8x1
105 x
0.0000
1.4373
0.0000
0.0013
0.0000
0.0004
0.0215
0.0017
```

Next Steps: How do I generalize to multiple data files?

Introducing **datastore** for handling multiple data files from multiple locations across multiple formats.

datastore is a container datatype that points to where data lives and stores information about how it should be read in.

```
ds = datastore('Experimental_Data\*.csv');
ds.VariableNames =
{'NegControl', 'Conc1', 'Conc2', 'Conc3', 'Conc4', 'Conc5', 'Conc6', 'Conc7', 'Conc8', 'Conc9',
'Conc10', 'PosControl'};
ds.ReadSize = 'file'
```

```
ds =
    TabularTextDatastore with properties:
```

```
Files: {
```

```

        ' ...\MicrotiterPlateFittingDemo\Experimental_Data\microtiter_data0001.csv';
        ' ...\MicrotiterPlateFittingDemo\Experimental_Data\microtiter_data0002.csv';
        ' ...\MicrotiterPlateFittingDemo\Experimental_Data\microtiter_data0003.csv'
        ... and 27 more
    }
    Folders: {
        ' ...\MicrotiterPlateFittingDemo\Experimental_Data'
    }
    FileEncoding: 'UTF-8'
    AlternateFileSystemRoots: {}
    VariableNamingRule: 'modify'
    ReadVariableNames: false
    VariableNames: {'NegControl', 'Conc1', 'Conc2' ... and 9 more}
    DatetimeLocale: en_US

Text Format Properties:
    NumHeaderLines: 0
    Delimiter: ','
    RowDelimiter: '\r\n'
    TreatAsMissing: ''
    MissingValue: NaN

Advanced Text Format Properties:
    TextscanFormats: {'%f', '%f', '%f' ... and 9 more}
    TextType: 'char'
    ExponentCharacters: 'eEdD'
    CommentStyle: ''
    Whitespace: ' \b\t'
    MultipleDelimitersAsOne: false

Properties that control the table returned by preview, read, readall:
    SelectedVariableNames: {'NegControl', 'Conc1', 'Conc2' ... and 9 more}
    SelectedFormats: {'%f', '%f', '%f' ... and 9 more}
    ReadSize: 'file'
    OutputType: 'table'
    RowTimes: []

Write-specific Properties:
    SupportedOutputFormats: ["txt"      "csv"      "xlsx"      "xls"      "parquet"      "parq"]
    DefaultOutputFormat: "txt"

```

Now we can perform the fitting on the preprocessed data. Here we initialize an empty array (**results**) to capture the EC50's and operate on the datastore until there is no more data left to read.

*NOTE: within the **for** loop, MATLAB Code Analyzer has identified that the variable, **results**, is [increasing in size with every iteration](#). This is not a recommended practice and should be avoided when possible.

```

results = [];

while hasdata(ds)
    data = read(ds);

    data.Variables = data.Variables - mean(data.NegControl);
    data.Variables = data.Variables ./ data.PosControl*100;

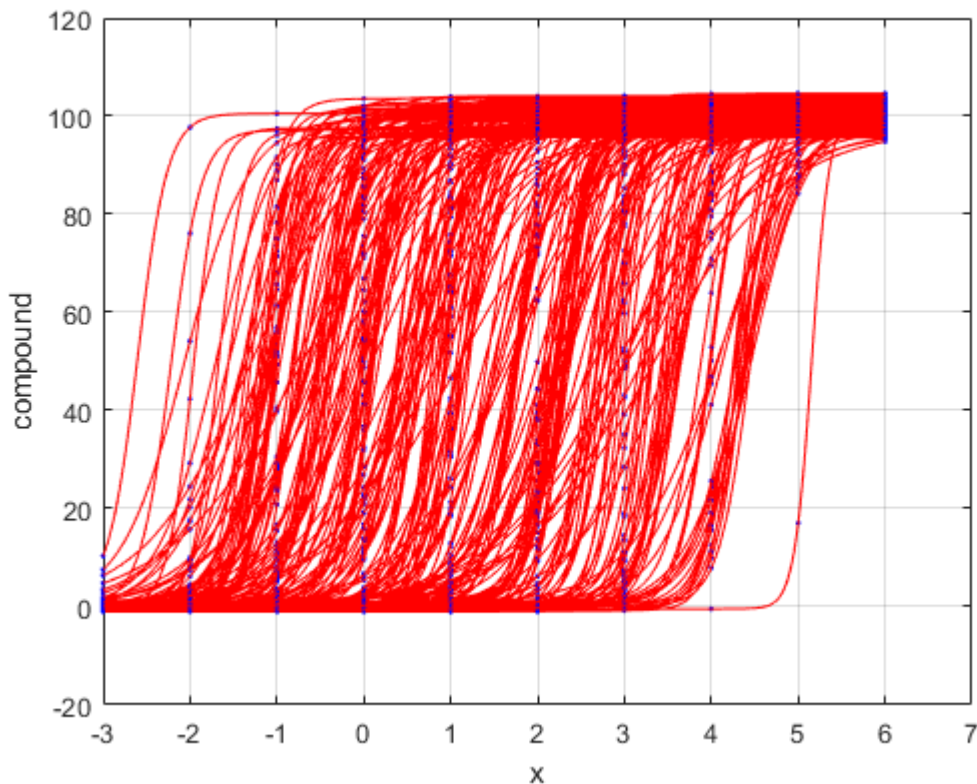
    data = removevars(data, 'NegControl');
    data = removevars(data, 'PosControl');

```

```

h = height(data);
for ii = 1:h
    compound = data{ii,1:10};
    [fitresult, gof] = createMicrotiterFit(x, compound);
    results = [results;fitresult.EC50];
end
end

```



```

writematrix(results,'myresults.csv')
reset(ds)

```

Extension: Alternatives for handling big data or scaling up

What if all the data needs to be processed at once? Is there any faster way of performing this analysis? There are several alternatives to explore...

```

do = false;

if do
    edit BigDataAndTall.mlx
end

```